

REVIEW

Pimpinella Species (Anise): Traditional Use, Mineral, Nutrient and Chemical Contents, Biological Activities

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Abstract

Plants, which are natural materials of great importance to humans, are currently utilized for various purposes. The use of plants with nourishing properties for treating diseases has a long history. This study compiled the general characteristics, usage areas, mineral and nutrient contents, biological activities, and chemical compositions of the *Pimpinella* species reported in the literature. The literature review reveals that the plant is widely used in traditional medicine. Furthermore, it has been determined that they may serve as significant natural resources regarding their mineral, nutritional, and chemical content. Furthermore, it has been observed, based on reported data in the literature, that *Pimpinella* species exhibit high levels of antioxidant and antimicrobial activity. Within this context, it has been observed that *Pimpinella* species can be significant sources not only in terms of their nutritional properties but also from a medicinal perspective.

Keywords: Antimicrobial, Anise, Antioxidant, Medicinal Plants, Traditional Medicine.

INTRODUCTION

Various natural materials, such as fungi, animals, and plants, are utilized in traditional medicine.¹ Among these natural products, plants are significant materials. Plants are utilized for various purposes such as food, spices, combating diseases, heating, and shelter in different cultures.² Plants constitute significant components of many human diets. They possess highly nutritious properties because they contain vitamins, minerals, and essential nutrients.³ Plants that stand out with their nourishing properties are also significant materials from a medical perspective.^{4,5} Numerous studies have reported that plants possess various activities such as antioxidant, anticancer, antimicrobial, anti-inflammatory, hepatoprotective, anti-aging, anti-allergic, and DNA protective properties.⁶⁻¹³ Determining the biological potential of plants is of great importance within this context. This study compiled the reported usage areas, general characteristics, nutritional and mineral

contents, biological activities, and chemical compounds of *Pimpinella* species from the literature.

GENUS *PIMPINELLA* AND USAGE AREAS

Pimpinella species belonging to the Apiaceae family are plants with a distribution range of 150 species across Europe, Asia, and Africa. The types refer to annual and perennial plants. From a morphological perspective, the plant possesses cordate-ovate or rectangular-ovate leaves and fruits that are slightly compressed laterally, each with five filiform veins.¹⁴⁻¹⁸

Pimpinella species are favored in alternative medicine in countries such as China, Egypt, Iran, Lebanon, and Palestine. It has been reported that there is still usage even in countries such as England and Italy, where the use of herbal medicines is low. Türkiye is the country where *Pimpinella* is most commonly utilized and preferred. The species of *Pimpinella* are commonly preferred for their above-

ground, root, and seed parts. The above-ground parts of the plant are utilized for their medicinal properties, including their ability to alleviate gastrointestinal distress, bronchial asthma, insomnia, persistent cough, renal colic, flatulence, expectorant, sedative, antidepressant, antiseptic, antispasmodic, analgesic, diuretic, estrogenic, and pectoral stimulant effects.

Table 1. Mineral and Nutritional Contents of *Pimpinella* species.

Nutritional Composition	Values (%)
Protein	13.35-28.73 %
Lipids	2.66-9.02 %
Carbohydrate	9.87-61.09 %
Crude fibers	2.83-33.50 %
Moisture	4.9-80.42 %
Ash	5.69-19.14 %
Mg	6.45-478.6 mg/kg
K	93.13-6332 mg/kg
Na	5.44- 365.10 mg/kg
Fe	0.15-1512 mg/kg
Ca	12.64-3141 mg/kg
Zn	0.12-14.31 mg/kg
Cu	0.029-0.035 mg/kg

The seed components exhibit various pharmacological properties such as abdominal pain

relief, gastrointestinal disorder management, carminative effects, expectorant properties, sedative effects, antidepressant properties, and antiseptic properties. Insomnia, persistent cough, carminative, stomach upset, stomach ache, calming, colic, tranquilizer, diuretic, flu, and various alcoholic beverages such as "patis" and "sambuca" are commonly used for their therapeutic properties as hypnotics, antispasmodics, expectorants, and in the treatment of epilepsy. The root parts, on the other hand, are utilized in the treatment of asthma, bronchitis, and menstrual disorders.¹⁹⁻²⁶

NUTRITIONAL AND MINERAL CONTENTS

Plants are an essential component of human diets and cannot be substituted. The food products that many individuals prefer to consume during each meal are commonly observed.²⁷ In this study, the nutritional and mineral contents of *Pimpinella* species reported in the literature are presented in Table 1.

BIOLOGICAL ACTIVITY

In the literature, various extracts such as ethanol, water, acetone, methanol, essential oil, aqueous, hexane, methylene chloride, hydroalcoholic, benzene, ethyl acetate, and n-butanol have been reported to be utilised *in vitro* and *in vivo* biological activity studies on *Pimpinella* species. The biological activity studies conducted on *Pimpinella* species reported in the literature are presented in Table 2.

Table 2. Biological activity of *Pimpinella* species.^{32, 35-54}

Plant species	Biological activities	Extractions
<i>P. anisoides</i> V.Brigr.	Antioxidant, anti-inflammatory	Ethanol
<i>P. anisum</i> L.	Antioxidant, antimicrobial, cytotoxicity, antiproliferative, anticancer, antispasmodic, antidiabetic, antiviral, insecticidal	Ethanol, water, methanol, essential oil, aqueous, hexane, methylene chloride, hydroalcoholicbenzene, ethylacetate, n-butanol
<i>P. aurea</i> DC.	Antioxidant, antimicrobial	Methanol
<i>P. brachycarpa</i> (Kom.) Nakai	Antioxidant, antimicrobial	Ethanol, methylene chloride, ethylacetate, hexane, methanol
<i>P. candolleana</i> Wight & Arn.	Antioxidant, α -glucosidase inhibitory	Ethanol, methanol
<i>P. puberula</i> (DC.) Boiss.	Antimicrobial	Essential oil
<i>P. saxifraga</i> L.	Antioxidant, antimicrobial	Essential oil
<i>P. stewartii</i> Nasir	Antioxidant, acetylcholinesterase	Acetone, water, aqueous, ethanol, ethylacetate
<i>P. thellungiana</i> H. Wolff	Antioxidant	Essential oil

Antioxidant activity

Living organisms synthesize numerous compounds with oxidizing properties due to their metabolic activities. While compounds of this particular book do not exhibit harmful effects at low levels, they can cause cellular damage as their levels increase.⁵⁵ The antioxidant defense system plays a role in suppressing the formation of oxidizing compounds, such as reactive oxygen species, in the presence of oxidizing agents. In cases where the antioxidant defense system is insufficient, oxidative stress occurs.⁵⁶ Numerous diseases such as Alzheimer's, Parkinson's, multiple sclerosis, cancer, and cardiovascular disorders may manifest due to oxidative stress.^{57,58} Supplementation with antioxidants can be utilized to reduce the effects of oxidative stress.⁵⁹ Plants are considered significant natural sources of supplementary antioxidants.⁶⁰

The literature reports the values of protein (13.35-28.73%), lipids (2.66-9.02%), carbohydrate (9.87-61.09%), crude fibers (2.83-33.50%), moisture (4.9-80.42%), and ash (5.69-19.14%) for *Pimpinella* species.²⁸⁻³² Furthermore, it has been reported that *Pimpinella* species contain Mg (6.45-478.6 mg/kg), K (93.13-6332 mg/kg), Na (5.44-365.10 mg/kg), Fe (0.15-1512 mg/kg), Ca (12.64-3141 mg/kg), Zn (0.12-14.31 mg/kg), and Cu (0.029-0.035 mg/kg).^{30,32-34} Within this context, it is believed that *Pimpinella* species may serve as a natural source of nutrients and minerals.

According to the literature, the antioxidant effect of the ethanol extract of *P. anisoides* collected from Italy was reported using the DPPH assay, with an LC50 value of 3.02 mg/mL.³⁵ The antioxidant activities of *P. anisum* extracts obtained from water and ethanol sourced from Türkiye were reported using various methods, including reducing power, superoxide anion scavenging, free radical scavenging, metal chelating, scavenging of hydrogen peroxide, and total antioxidant activity. According to the test results, it has been reported that plant extracts exhibit high antioxidant activities.³⁷ According to a study conducted in Egypt, the ethanol and aqueous extracts of *P. anisum*'s seed and aerial parts exhibited DPPH activities ranging from 13.7% to 91.3% at concentrations of 0.05-0.3 mg/mL.⁴⁶ It has been reported that the volatile oil of *P. anisum* collected from Greece exhibited antioxidant activity with DPPH and ABTS assay values of 48% and 18.6%, respectively.⁴⁴ According to reports, the IC50 values of the methanol, volatile oil, polar subfraction, and non-polar subfraction of *P. aurea* collected from Iran were found to be between 108-549 µg/mL in the

DPPH test. Furthermore, it has been reported that the LC values of β-carotene/linoleic acid test results vary between 5.98% and 65.87.⁴⁷ It has been reported that the ethanol extract of *P. brachycarpa* collected from South Korea exhibits a DPPH anion scavenging activity value was 8.80 mg/g, ABTS cation scavenging activity value was 63.53 mg/g, and a reducing power test result was 0.85.⁴⁹ According to reports, the IC50 value of *P. candolleana* extracts collected from China, including petroleum ether, methanol, and ethanol, varied between 25.46-27.20 µg/mL in the DPPH test, 9.23-21.69 µg/mL in the ABTS test, and 138.88-919.84 µmol/g in the FRAP test.⁵⁰ It has been reported that the IC50 value of the volatile oil of *P. saxifraga* collected from Tunisia for DPPH radical scavenging activity is 6.81 µg/mL, the IC50 value for β-carotene bleaching inhibition test is 206 µg/mL, the EC50 value for ferric reducing power test is 35.20 µg/mL, and the total antioxidant activity test result is 213.96 µmol/mL.⁵² The DPPH values of aqueous and acetone extracts of *P. stewartii* collected from Pakistan were reported as 61.08% and 62.39%, respectively, ferrous ion chelating values were reported as 61.16% and 50.76%, and hydrogen peroxide scavenging values were reported as 25.18 and 52.59%, respectively. In addition, the hydroxyl radical scavenging value was reported 0.091 and 11.70%, respectively, the phosphomolybdenum complex assay value was 56.55 and 86.26 µM/100 g, and the ferric ion reducing antioxidant power test result was 23.28 and 14.24 µM/100 g, respectively.³² According to a study conducted in Kazakhstan, the volatile oil obtained from the root and aerial parts of *P. thellungiana* exhibited anti-radical activity ranging from 2.9% to 12.7% at concentrations between 0.1-1 mg/mL.⁵⁴ Upon reviewing the literature on antioxidant activity studies conducted on *Pimpinella* species within this scope, it is observed that *Pimpinella* species exhibit high antioxidant properties. It is believed that *Pimpinella* species may serve as a natural source of antioxidants within this context.

Antimicrobial activity

There is an increasing trend in the incidence of diseases caused by microorganisms.⁶¹ The emergence of resistant microorganisms due to improper medication use renders the antimicrobial drugs used insufficiently. In this context, researchers are directing their attention toward novel antimicrobial sources.⁶² The potential side effects of synthetic drugs have led individuals to turn towards natural antimicrobial agents. Within this context, plants are significant sources of natural antimicrobial

agents.^{63,64} In this study, the antimicrobial potential of *Pimpinella* species was compiled based on literature data (Table 2). It has been reported that the water and ethanol extracts of *P. anisum* collected from Türkiye exhibit inhibition zones ranging from 7-11 mm against *Pseudomonas aeruginosa*, *Escherichia coli*, *Proteus mirabilis*, *Citrobacter koşeri*, *Enterobacter aerogenes*, *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Micrococcus luteus*, *Staphylococcus epidermidis*, and *Candida albicans*.³⁷ It has been reported that the volatile oil and methanol extracts of *P. anisum* collected from Iraq exhibited the most favorable outcomes against *Staphylococcus aureus*, *Bacillus cereus*, and *Proteus vulgaris* at a concentration of 62.5 µg/mL.³⁹ According to a study conducted in Egypt, ethanol and aqueous extracts of *P. anisum*'s seed and aerial parts exhibited inhibition zones ranging from 2.7-21 mm against *Bacillus cereus*, *Staphylococcus aureus*, *Salmonella typhimurium*, and *Escherichia coli* at concentrations ranging from 1.25-5 mg/mL.⁴⁶ It has been reported that silver nanoparticles produced using aqueous extracts of *P. anisum* collected from Saudi Arabia exhibit minimum and average diameter values ranging from 3.2 to 16 nm against *Acinetobacter baumannii*, *Klebsiella pneumoniae*, *S. typhi*, and *P. aeruginosa*.⁴³ According to the literature, *P. anisum* collected from Iraq effectively against *S. aureus*, *B. cereus*, and *E. coli* at a concentration of 31.2 µg/mL. Additionally, it was effective against *P. mirabilis* and *K. pneumonia* at a concentration of 62.5 µg/mL, against *Candida albicans* at a concentration of 500.0 µg/mL, and against *Pseudomonas aeruginosa* at a concentration greater than 500.0 µg/mL.⁴⁰ It has been reported that the volatile oil of *P. aurea* collected from Iran exhibits inhibition zones ranging from 10-16 mm against *K. pneumoniae*, *P. vulgaris*, *S. dysenteriae*, and *S. aureus*.⁴⁷ It has been reported that the ethanol extract of *P. brachycarpa* collected from South Korea exhibits antimicrobial effects against *S. aureus* and *B. subtilis*.⁴⁹ A study reported from South Korea indicates that *P. brachycarpa* has MIC and MFC values of 0.25 mg/ml and 0.5 mg/ml, respectively, against *Aspergillus niger*.⁴⁸ It has been reported that the inhibition zone values of *P. aeruginosa*, *Bacillus cereus*, *Micrococcus luteus*, *S. aureus*, *Yersinia enterocolitica*, and *C. albicans* vary between 3-36 mm when exposed to the volatile oil of *P. puberula* collected from Iran.⁵¹ It has been reported that the volatile oil of *P. saxifraga* collected from Tunisia has MIC values ranging from 0.78 to 3.125 mg/mL

against *E. coli*, *S. typhimurium*, *P. aeruginosa*, *B. cereus*, *M. luteus*, and *L. monocytogenes*.⁵² Upon examination of the studies reported in the literature regarding *Pimpinella* species, it is evident that these plants possess significant antimicrobial potential. Within this context, it is believed that *Pimpinella* species may serve as natural antimicrobial agents.

Other activities

The literature has reported that *Pimpinella* species exhibit various biological activities in addition to their antioxidant and antimicrobial activities (Table 2). The ethanol extract of *P. anisoides* collected from Italy was utilised to investigate its anti-inflammatory effect on murine monocytic macrophage cell line RAW 264.7. It has been reported that the working outcome possesses an IC50 value of 72.7 µg/mL *in vivo*.³⁵ It has been reported that silver nanoparticles produced using aqueous extracts of *P. anisum* collected from Saudi Arabia showed minimal negative effects on cell proliferation, without any changes in cell viability, when tested against a skin stromal cell line (hSSC) and a cancer cell line (HT115) at various concentrations of less than 10 µg. Furthermore, it has been reported to lead to increased cytotoxicity at doses exceeding 10 µg.⁴³ It has been reported that the EC50 values of the essential oil of *P. anisum* collected from Greece against HepG2, Caco2, MCF-7, and THP-1 cell lines were 0.39, 0.25, 0.3, and 0.11 mg/mL, respectively.⁴⁴ A study in Israel reported that the ethanol extract of *P. anisum* exhibited robust proliferation on rat skeletal muscle cell line (L6) and human prostate cancer cell line (PC-3) at an IC50 value of 400 µg/mL.⁴¹ The hydroalcoholic extract of *P. anisum* collected from Brazil was utilised to determine its antispasmodic effect on the anococcygeus smooth muscle of rats. According to the study findings, it has been reported that the extract used inhibited the contraction caused by acetylcholine at a concentration of 50 µg/mL.³⁸ In a study conducted in India, the IC50 values of the highest α-amylase and α-glucosidase effects of the ethyl acetate fraction of *P. anisum* were reported as 0.12 and 0.15 mg/mL, respectively.⁴² A study conducted in India reported that the volatile oil of *P. anisum* exhibited inhibitory effects against Potato virus X, Tobacco mosaic virus, and Tobacco ringspot virus at a concentration of 300 ppm.³⁶ It has been reported that the insecticidal effect of the essential oil of *P. anisum* collected from Egypt against the red flour beetle, *Tribolium castaneum*, has an LC50 value of 9.3% v/v.⁴⁵ It has been reported that the petroleum ether, methanol, and ethanol extracts of *P. candolleana* collected from China exhibit α-

glucosidase inhibitory activity with an LC50 value ranging from 4.42-68.71 µg/ml, attributed to the chemical compounds present in *P. candolleana*.⁵⁰ The IC50 values of the acetylcholinesterase activity of the water and ethyl acetate fractions of *P. stewartii* collected from Pakistan were reported as 72.6 and 30.41 µg/mL, respectively.⁵³

CHEMICAL CONTENTS

In plants, numerous biologically active compounds

are synthesised within their structures. Numerous studies have reported significant biological activities of compounds found in plants.⁶⁵ In this study, the chemical compounds identified in *Pimpinella* species in the literature have been compiled. Within this context, it has been observed that studies have been conducted on the aerial, stem, and seed parts of *Pimpinella* species in the literature. The findings obtained were presented in Table 3.

Table 3. Chemical contents of *Pimpinella* species.^{35, 51, 54, 66-78}

Plant species	Geographic regions	Used Parts	Chemical contents
<i>P. acuminata</i> (Edgew.) C.B. Clarke	India, Pakistan	Aerial, stem	β-caryophyllene (12.5%), dill apiole (11.3-20.4%), parsley apiole (39.9-61.8%), myristicin (16.2%), methyl coniine (70.0%), coniine (4.0%), n-pentadecane (4.0%), 1-methyl-2-pentyl piperidene (3.3%), heptadecane (3.0%), apiole (1.5%)
<i>P. anisoides</i> V.Brig.	Italy	Aerial	Trans-anethole (54.5%), limonene (13.5%), sabinene (4.4%)
<i>P. anisum</i> L.	Estonia, Serbia, Iran, Brazil, Algeria, Pakistan, Egypt	Aerial, seed	Trans-anethole (65.6–93.7%), γ-himachalene (0.4–8.2%), trans-pseudoisoeugenyl 2-methylbutyrate (0.4–6.4%), p-anisaldehyde (<1-5.4%), methylchavicol (0.5–2.3%), cis-isoeugenol (1.99%), linalool (1.79%), (E)-anethole (80.7-90.35%), estragole (1.9-5.6%), eugenyl acetate (3.34-3.92%), α-zingiberene (1.9%), cis-pseudoisoeugenyl 2-methylbutyrate (~3%), o-isoeugenol (1.9%), fenchone (5.6%), camphor (3.1%)
<i>P. aurea</i> DC.	Iran	Aerial, stem, seed	Limonene (8.9-21.4%), viridiflorol (12.8-37.0%), α-pinene (11.5%), kessane (10.5%), germacrene D (4.9%), β-bisabolene (4.2-50.8%), α-zingiberene (3.3%), citronellyl acetate (3.1%), caryophyllene oxide (6.6%), 1,8-cineol (8.9-21.4%), estragol (5.1%), trans-a-bergamotene (72.8%)
<i>P. puberula</i> (DC.) Boiss.	Iran	Aerial, stem, seed	Limonene (21.7-82.4%), pregejerene (14.6-55.4%), geijerene (7.2-11.7%)
<i>P. saxifraga</i> L.	Iran	Aerial	Trans-α-bergamotene (20.1%), β-sesquiphellandrene (10.8%), β-bisabolene (10.1%)
<i>P. thellungiana</i> H. Wolff	Kazakhstan	Aerial, stem	Hexenal (0.2-8.9%), β-pinene (0.4-4.1%), undecane (1.5-15.6%), geijerene (0.6-9.2%), 9-Methyl-10-methylenetricyclo[4.2.1.1(2,5)]decan-9-ol (3.4%), cis-β-farnesene (0.3-12.6%), β-bisabolene (1.6-18.8%), spathulenol (3.3%), caryophyllene oxide (0.7-4.3%), 2-methyl-4-methoxy-2-(3-methoxyoxiranyl)phenylbutanoate (0.4-29.5%)
<i>P. tragiun</i> Vill.	Iran	Aerial, stem	Germacrene D (6.2-34.7%), germacrene B (14.1-18.3%), bornyl acetate (4.1-15.8%), β-caryophyllene (4.8-7.3%) β-pinene (4.5-25.3%), hexadecanol (4.7-10.3%), sabinene (13.6%)

In the literature, the main components in the essential oil content of *P. acuminata* species are β-caryophyllene (12.5%), dill apiole (11.3-20.4%), parsley apiole (39.9-61.8%), myristicin (16.2%), methyl coniine (70.0%), Coniine (4.0%), n-pentadecane (4.0%), 1-methyl-2-pentyl piperidene (3.3%), heptadecane (3.0%), and apiole (1.5%) have been reported.^{66,67} It has been reported that the main components in the essential oil content of *P. anisoides* species are trans-anethole (54.5%),

limonene (13.5%) and sabinene (4.4%).³⁵ It has been reported that the main components in the essential oil content of *P. anisum* species are trans-anethole (65.6–93.7%), γ-himachalene (0.4–8.2%), trans-pseudoisoeugenyl 2-methylbutyrate (0.4–6.4%), p-anisaldehyde (<1-5.4%), methylchavicol (0.5–2.3%), cis-isoeugenol (1.99%), linalool (1.79%), (E)-anethole (80.7-90.35%), estragole (1.9-5.6%), eugenyl acetate (3.34-3.92%), α-zingiberene (1.9%), cis-pseudoisoeugenyl 2-methylbutyrate (~3%), o-

isoeugenol (1.9%), fenchone (5.6%) and camphor (3.1%).⁶⁸⁻⁷⁴ It has been reported that the main components in the essential oil content of *P. aurea* species are limonene (8.9-21.4%), viridifluorool (12.8-37.0%), α -pinene (11.5%), chestnut (10.5%), germacrene D (4.9%), β -bisabolene (4.2-50.8%), α -zingiberene (3.3%), citronellyl acetate (3.1%), caryophyllene oxide (6.6%), 1,8-cineol (8.9-21.4%), estragole (5.1%) and trans- α -bergamotene (72.8%).^{75,76} It has been reported that the main components in the essential oil content of *P. puberula* species are limonene (21.7-82.4%), pregeijerene (14.6-55.4%) and geijerene (7.2-11.7%).⁵¹ It has been reported that the main components in the essential oil content of *P. saxifraga* species are trans- α -bergamotene (20.1%), β -sesquiphellandrene (10.8%) and β -bisabolene (10.1%).⁷⁷ It has been reported that the main components in the essential oil content of *P. thellungiana* species are hexenal (0.2-8.9%), β -pinene (0.4-4.1%), undecane (1.5-15.6%), geijerene (0.6-9.2%), 9-Methyl-10-methylenetricyclo [4.2.1.1 (2.5)]decan-9-ol (3.4 %), cis - β -farnesene (0.3-12.6%), β -bisabolene (1.6-18.8%), spathulenol (3.3%), caryophyllene oxide (0.7-4.3%) and 2-methyl-4-methoxy-2-(3-methoxyoxiranyl) phenylbutanoate (0.4-29.5%).⁵⁴ (Suleimen et al., 2017). It has been reported that the main components in the essential oil content of *P. tragium* species are hexadecanol (4.7-10.3%), germacrene D (6.2-34.7%), germacrene B (14.1-18.3%), bornyl acetate (4.1-15.8%), β -caryophyllene (4.8-7.3%) β -pinene (4.5-25.3%), and sabinene (13.6%).⁷⁸ In this context, based on the literature data, it has been reported that the 5 highest reported compounds in *Pimpinella*

species are trans- α -bergamotene (72.8%), methylconiine (70.0%), trans-anethole (65.6–93.7%), (E)-anethole (80.7-90.35%), and limonene (21.7-82.4%). In this context, it is thought that *Pimpinella* species may be a natural source for the compounds reported in its body.

CONCLUSION

This study compiles the literature on *Pimpinella* species reported in previous studies. Within this scope, the general characteristics, mineral and nutrient contents, areas of use, biological activities, and chemical compositions have been compiled. Based on the literature review conducted, it has been determined that *Pimpinella* species are commonly used in traditional medicine. Furthermore, it is believed that the reported minerals, nutrients, and chemical contents within it could potentially serve as a natural resource. Additionally, it has been reported in the literature that *Pimpinella* species exhibit high antioxidant and antimicrobial activities. As a result, it has been observed that in addition to the nutritious properties of *Pimpinella* species, they could be significant natural materials in pharmacological designs in future studies.

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REFERENCES

1. Mohammed FS, Sevindik M, Uysal I, Sevindik E, Akgül H. A Natural Material for Suppressing the effects of oxidative stress: biological activities of *Alcea kurdica*. *Biol Bull.* 2022; 49(Suppl 2): S59-S66.
2. Sevindik M, Akgul H, Pehlivan M, Selamoglu, Z. Determination of therapeutic potential of *Mentha longifolia ssp. longifolia*. *Fresen Environ Bull.* 2017;26(7):4757-4763.
3. Mohammed FS, Günel S, Şabik AE, Akgül H, Sevindik M. Antioxidant and Antimicrobial activity of *Scorzonera papposa* collected from Iraq and Turkey. *KSÜ Tar Doga Derg.* 2020a;23(5): 1114-1118.
4. Mohammed FS, Günel S, Pehlivan M, Doğan M, Sevindik M, Akgül H. Phenolic content, antioxidant and antimicrobial potential of endemic *Ferulago platycarpa*. *Gazi Univ J Sci.* 2020b;33(4): 670-677.
5. Korkmaz N, Dayangaç A, Sevindik M. Antioxidant, antimicrobial and antiproliferative activities of *Galium aparine*. *J Fac Pharm Ankara.* 2021;45(3): 554-564.
6. Tewtrakul S, Subhadhirasakul S. Anti-allergic activity of some selected plants in the Zingiberaceae family. *J Ethnopharmacol.* 2007;109(3): 535-538.
7. Hassan A, Rahman S, Deeba F, Mahmud S. Antimicrobial activity of some plant extracts having hepatoprotective effects. *J Med Plant Res.* 2009;3(1): 20-023.
8. Yasin ZA, Ibrahim F, Rashid NN, Razif MF, Yusof R. The importance of some plant extracts as skin anti-aging resources: a review. *Curr Pharm Biotechnol.* 2017;18(11): 864-876.
9. Mohammed FS, Akgul H, Sevindik M, Khaled BMT. Phenolic content and biological activities of *Rhus coriaria var. zebaria*. *Fresen Environ Bull.* 2018;27(8): 5694-5702.

10. Mohammed FS, Karakaş M, Akgül H, Sevindik M. Medicinal properties of *Allium calocephalum* collected from Gara Mountain (Iraq). *Fresen Environ Bull.* 2019;28(10):7419-7426.
11. Mohammed FS, Pehlivan M, Sevindik E, Akgul H, Sevindik M, Bozgeyik I, Yumrutas O. Pharmacological properties of edible *Asparagus acutifolius* and *Asparagus officinalis* collected from North Iraq and Turkey (Hatay). *Acta Aliment.* 2021a;50(1): 136-143.
12. Unal O, Eraslan EC, Uysal I, Mohammed FS, Sevindik M, Akgul H. Biological activities and phenolic contents of *Rumex scutatus* collected from Turkey. *Fresen Environ Bull.* 2022;31(7):7341-7346.
13. Uysal I, Koçer O, Mohammed FS, Lekesiz Ö, Doğan M, Şabik AE, Sevindik E, Gerçeker FO, Sevindik M. Pharmacological and Nutritional Properties: Genus *Salvia*. *Adv Pharmacol.* 2023;11(2): 140-155
14. Pimenov MG, Leonov MV. The Asian Umbelliferae biodiversity database (ASIUM) with particular reference to South-West Asian taxa. *Turkish J. Bot.* 2004; 28(1): 139-145.
15. Pu FT, Watson MF. *Pimpinella* L. Flora of China, 2005;14: 93-104.
16. Spalik K, Downie SR. Intercontinental disjunctions in *Cryptotaenia* (Apiaceae, Oenantheae): an appraisal using molecular data. *J Biogeogr.* 2007;34(12): 2039-2054.
17. Magee AR, van Wyk BE, Tilney PM, Downie SR. Phylogenetic position of African and Malagasy *Pimpinella* species and related genera (Apiaceae, Pimpinelleae). *Plant Syst. Evol.* 2010;288(3): 201-211.
18. Fernández-Prieto JA, Sanna M, Bueno Sánchez Á, Molero-Mesa J, Llorens García L, Cires E. Polyphyletic origin in *Pimpinella* (Apiaceae): evidence in Western Europe. *J. Plant Res.* 2018;131(5): 747-758.
19. AbouZid SF, Mohamed AA. Survey on medicinal plants and spices used in Beni-Sueif, Upper Egypt. *J Ethnobiol Ethnomedicine* 2011;7(1): 1-6.
20. Hammer K, Laghetti G, Cifarelli S, Spahillari M, Perrino P. *Pimpinella anisoides* Briganti. *Genet. Resour. Crop Evol.* 2000;47(2): 223-225.
21. Kreydiyyeh SI, Usta J, Knio K, Markossian S, Dagher S. Aniseed oil increases glucose absorption and reduces urine output in the rat. *Life Sci.* 2003;74(5): 663-673.
22. Yöney A, Prieto JM, Lardos A, Heinrich M. Ethnopharmacy of turkish-speaking cypriots in greater London. *Phytother Res.* 2010; 24(5): 731-740.
23. Kang Y, Luczaj L, Ye S, Zhang S, Kang J. Wild food plants and wild edible fungi of Heihe valley (Qinling Mountains, Shaanxi, central China): herbophilia and indifference to fruits and mushrooms. *Acta Soc Bot Pol.* 2012;81(4): 405-413.
24. Tetik F, Civelek S, Cakilcioglu U. Traditional uses of some medicinal plants in Malatya (Turkey). *J Ethnopharmacol.* 2013;146(1): 331-346.
25. Tepe AS, Tepe B. Traditional use, biological activity potential and toxicity of *Pimpinella* species. *Ind Crops Prod.* 2015; 69: 153-166.
26. Sun W, Shahrajabian MH, Cheng Q. Anise (*Pimpinella anisum* L.), a dominant spice and traditional medicinal herb for both food and medicinal purposes. *Cogent Biol.* 2019;5(1): 1673688.
27. Mohammed FS, Korkmaz N, Doğan M, Şabik AE, Sevindik M. Some medicinal properties of *Glycyrrhiza glabra* (Licorice). *J Fac Pharm Ankara.* 2021b;45(3): 524-534.
28. Ciftci M, Guler T, Dalkiliç B, Ertas ON. The effect of anise oil (*Pimpinella anisum* L.) on broiler performance. *Int J Poult Sci.* 2005;4(11): 851-855.
29. Lee JJ, Choo MH, Lee MY. Physicochemical compositions of *Pimpinella brachycarpa*. *J Korean Soc Food Sci Nutr.* 2007;36(3): 327-331.
30. Chae HS, Lee SH, Jeong HS, Kim WJ. Antioxidant activity and physicochemical characteristics of *Pimpinella brachycarpa* Nakai with treatments methods. *Korean J Food Nutr.* 2013;26(1): 125-131.
31. Hassan FAS, Ali EFA. Comparative Study between Traditional Mineral Nutrition and Alternative Sources on Anise *Plant Eur J Sci Res.* 2013;106(2): 201-212.
32. Abbasi AM, Xinbo-Guo X. Proximate composition, phenolic contents and in vitro antioxidant properties of *Pimpinella stewartii* (A wild medicinal food). *J. Food Nutr Res.* 2015;3(5): 330-336
33. Kumaravel S, Alagusundaram K. Determination of mineral content in Indian spices by ICP-OES. *Orient J Chem.* 2014;30(2):631-636.
34. Bae YJ, Kim MH, Lee JH, Choi MK. Analysis of six elements (Ca, Mg, Fe, Zn, Cu, and Mn) in several wild vegetables and evaluation of their intakes based on Korea National Health and Nutrition Examination Survey 2010–2011. *Biol Trace Elem Res.* 2015;64(1): 114-121.
35. Conforti F, Tundis R, Marrelli M, Menichini F, Statti GA, De Cindio B, Houghton PJ. Protective effect of *Pimpinella anisoides* ethanolic extract and its constituents on oxidative damage and its inhibition of nitric oxide in lipopolysaccharide-stimulated RAW 264.7 macrophages. *J Med Food.* 2010;13(1): 137-141.
36. Shukla HS, Dubey P, Chaturvedi RV. Antiviral properties of essential oils of *Foeniculum vulgare* and *Pimpinella anisum* L. *Agronomie.* 1989;9(3): 277-279.
37. Gülçın İ, Oktay M, Kırçacı E, Küfrevioğlu Öİ. Screening of antioxidant and antimicrobial activities of anise (*Pimpinella anisum* L.) seed extracts. *Food Chem.* 2003;83(3): 371-382.

38. Tirapelli CR, de Andrade CR, Cassano AO, De Souza FA, Ambrosio SR, da Costa FB, de Oliveira AM. Antispasmodic and relaxant effects of the hidroalcoholic extract of *Pimpinella anisum* (Apiaceae) on rat anococcygeus smooth muscle. *J. Ethnopharmacol.* 2007;110(1): 23-29.
39. Al-Bayati FA. Synergistic antibacterial activity between *Thymus vulgaris* and *Pimpinella anisum* essential oils and methanol extracts. *J. Ethnopharmacol.* 2008;116(3): 403-406.
40. Mohammed MJ. Isolation and identification of anethole from *Pimpinella anisum* L. fruit oil. An antimicrobial study. *J Pharma Res.* 2009;2(5): 915-919.
41. Kadan S, Rayan M, Rayan A. Anticancer activity of anise (*Pimpinella anisum* L.) seed extract. *The Open Nutra J.* 2013;6(1): 1-5.
42. Shobha RI, Rajeshwari CU, Andallu B. Anti-peroxidative and anti-diabetic activities of aniseeds (*Pimpinella anisum* L.) and identification of bioactive compounds. *Am J Phytomedicine Clin Ther.* 2013;1(5): 516-527.
43. AlSalhi MS, Devanesan S, Alfuraydi AA, Vishnubalaji R, Munusamy MA, Murugan K, Benelli G. Green synthesis of silver nanoparticles using *Pimpinella anisum* seeds: antimicrobial activity and cytotoxicity on human neonatal skin stromal cells and colon cancer cells. *Int J Nanomedicine.* 2016;11, 4439.
44. Fitsiou E, Mitropoulou G, Spyridopoulou K, Tiptiri-Kourpeti A, Vamvakias M, Bardouki H, Pappa A. Phytochemical profile and evaluation of the biological activities of essential oils derived from the Greek aromatic plant species *Ocimum basilicum*, *Mentha spicata*, *Pimpinella anisum* and *Fortunella margarita*. *Molecules.* 2016;21(8): 1069.
45. Hashem AS, Awadalla SS, Zayed GM, Maggi F, Benelli G. *Pimpinella anisum* essential oil nanoemulsions against *Tribolium castaneum*—insecticidal activity and mode of action. *Environ Sci Pollut Res.* 2018;25(19): 18802-18812.
46. Amer AM, Aly UI. Antioxidant and antibacterial properties of anise (*Pimpinella anisum* L.). *Egypt Pharm J.* 2019;18(1): 68.
47. Safaei-Ghomi J, Ebrahimabadi AH, Djafari-Bidgoli Z, Kashi JF, Batooli H. Bioactive properties of oil and methanol extracts of *Pimpinella aurea* DC. *Am-Eurasian J Sustain Agric.* 2009;3(2): 151-156.
48. Ahn SM, Choi TH, Kwun IS, Sohn HY. Antifungal activity of methylene chloride fraction of *Pimpinella brachycarpa* against *Aspergillus niger*. *Microbiol Biotechnol Lett.* 2011;39(2):168-174.
49. Kim SJ, Cho AR, Han J. Antioxidant and antimicrobial activities of leafy green vegetable extracts and their applications to meat product preservation. *Food control.* 2013;29(1): 112-120.
50. Chang X, Kang W. Antioxidant and α -glucosidase inhibitory compounds from *Pimpinella candolleana* Wight et Arn. *Med Chem Res.* 2012;21(12): 4324-4329.
51. Askari F, Sefidkon F, Teymouri M, Yousef NS. Chemical composition and antimicrobial activity of the essential oil of *Pimpinella puberula* (DC.) Boiss. *J Agr Sci Tech.* 2009c;11: 431-438.
52. Ksouda G, Sellimi S, Merlier F, Falcimaigne-Cordin A, Thomasset B, Nasri M, Hajji M. Composition, antibacterial and antioxidant activities of *Pimpinella saxifraga* essential oil and application to cheese preservation as coating additive. *Food Chem.* 2019;288: 47-56.
53. Sahibzada MUK, Ahmad W, El-Shehawi AM, Jafri I, Irfan S, Khusro A, Emran TB. Acetylcholinesterase inhibitory activity and iron-induced lipid peroxidation reducing potential of *Pimpinella stewartii* leaves in male wistar rats. *J King Saud Univ Sci.* 2022;34(4): 101993.
54. Suleimen EM, Ibataev ZA, Iskakova ZB, Dudkin RV, Gorovoi PG, Aistova EV. Constituent composition and biological activity of essential oil from *Pimpinella thellungiana*. *Chem Nat Compd.* 2017;53(1): 169-172.
55. Krupodorova T, Sevindik M. Antioxidant potential and some mineral contents of wild edible mushroom *Ramaria stricta*. *AgroLife Sci J.* 2020;9(1): 186-191.
56. Bal C, Sevindik M, Akgul H, Selamoglu Z. Oxidative stress index and antioxidant capacity of *Lepista nuda* collected from Gaziantep/Turkey. *Sigma.* 2019;37(1): 1-5.
57. Selamoglu Z, Sevindik M, Bal C, Ozaltun B, Sen İ, Pasdaran A. Antioxidant, antimicrobial and DNA protection activities of phenolic content of *Tricholoma virgatum* (Fr.) P. Kumm. *Biointerface Res Appl Chem.* 2020;10(3):5500-5506
58. Saridogan BGO, Islek C, Baba H, Akata I, Sevindik M. Antioxidant antimicrobial oxidant and elements contents of *Xylaria polymorpha* and *X. hypoxylon* (Xylariaceae). *Fresenius Envir Bull.* 2021;30(5): 5400-5404.
59. Eraslan EC, Altuntas D, Baba H, Bal C, Akgül H, Akata I, Sevindik M. Some biological activities and element contents of ethanol extract of wild edible mushroom *Morchella esculenta*. *Sigma.* 2021;39(1): 24-28.
60. Uysal İ, Mohammed FS, Şabik AE, Kına E, Sevindik M. Antioxidant and Oxidant status of medicinal plant *Echium italicum* collected from different regions. *Turkish JAF Sci Tech.* 2021;9(10):1902-1904.
61. Baba H, Sevindik M, Dogan M, Akgül H. Antioxidant, antimicrobial activities and heavy metal contents of some Myxomycetes. *Fresen Environ Bull.* 2020;29(09):7840-7846.
62. Mohammed FS, Uysal I, Sevindik M. A review on antiviral plants effective against different virus types. *Prospects in Pharma Sci.* 2023; 21(2): 1-21.
63. Bal C, Akgul H, Sevindik M, Akata I, Yumrutas O. Determination of the anti-oxidative activities of six mushrooms. *Fresenius Environ Bull.* 2017; 26(10): 6246-6252.
64. Islek C, Saridogan BGO, Sevindik M, Akata I. Biological activities and heavy metal contents of some *Pholiota* species. *Fresen Environ Bull.* 2021;30(6): 6109-6114.

65. Akgul H, Korkmaz N, Dayangaç A, Sevindik M. Antioxidant potential of endemic *Salvia absconditiflora*. *Turkish JAF Sci Tech*. 2020;8(10): 2222-2224.
66. Ashraf M, Ahmad R, Mahmood S, Bhatti MK. Studies on the essential oils of the Pakistani species of the family Umbelliferae. Part 45. *Ferula assafoetida*, Linn (Herra Hing) gum oil. *Pak J Sci Ind Res*. 1980;23(1-2): 68-69.
67. Melkani AB, Javed MS, Melkani KB, Dev V, Beauchamp PS. Terpenoid composition of the essential oil from *Pimpinella acuminata* (Edgew.) CB Clarke. *J Essent Oil Res*. 2006;18(3): 312-314.
68. Omidbaigi R, Hadjiakhoondi A, Saharkhiz M. Changes in content and chemical composition of *Pimpinella anisum* oil at various harvest time. *J. Essent. Oil-Bear Pl*. 2003;6(1): 46-50.
69. Rodrigues VM, Rosa PT, Marques MO, Petenate AJ, Meireles MAA. Supercritical extraction of essential oil from aniseed (*Pimpinella anisum* L) using CO₂: solubility, kinetics, and composition data. *J Agric Food Chem*. 2003;51(6): 1518-1523.
70. Orav A, Raal A, Arak E. Essential oil composition of *Pimpinella anisum* L. fruits from various European countries. *Nat Prod Res*. 2008; 22(3):227-232.
71. Samojlik I, Mijatović V, Petković S, Škrbić B, Božin B. The influence of essential oil of aniseed (*Pimpinella anisum*, L.) on drug effects on the central nervous system. *Fitoterapia*. 2012;83(8): 1466-1473.
72. Saibi, S., Belhadj, M., Benyoussef, EH. (2012). Essential oil composition of *Pimpinella anisum* from Algeria. *Anal Chem Lett*. 2012;2(6): 401-404.
73. Ullah H, Mahmood A, Honermeier B. Essential oil and composition of anise (*Pimpinella anisum* L.) with varying seed rates and row spacing. *Pak J Bot*. 2014;46(5): 1859-1864.
74. Khalid AK. Quality and quantity of *Pimpinella anisum* L. essential oil treated with macro and micronutrients under desert conditions. *Int Food Res J*. 2015;22(6): 2396.
75. Askari F, Sefidkon F, Mozafarian V. Essential oil composition of *Pimpinella aurea* DC from Iran. *Flavour Fragr J*. 2005a;20(2): 115-117.
76. Assadian F, Masoudi S, Nematollahi F, Rustaiyan A, Larijani K, Mazloomifar H. Volatile constituents of *Xanthogalum purpurascens* Ave-Lall., *Eryngium caeruleum* MB and *Pimpinella aurea* DC. Three Umbelliferae herbs growing in Iran. *J Essent Oil Res*. 2005;17(3): 243-245.
77. Masoudi S, Rustaiyan A, Mazloomifar H. Composition of the Essential Oils of *Pimpinella anisactis* Rech. f. and *Pimpinella saxifraga* L. from Iran. *J Essent Oil Res*. 2009;21(2):146-148.
78. Askari F, Sefidkon F. Volatile components of *Pimpinella tragiun* Vill. from Iran. *Iran J Pharm Sci*. 2005b;2:117-120.