

Usnic acid uses mitochondrial apoptotic pathway in it's antitumoral role

Usnik asitin tümör önleyici rolü mitokondrial apoptotik yolak üzerindedir

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SUMMARY

Objective: Usnic acid is a secondary metabolite in lichens whose role has not been completely elucidated. Lichen extracts containing usnic acid have been utilized in medicine, perfumery, cosmetics, and ecology. Usnic asit inhibits cell growth, induces the cell cycle arrest and apoptosis in human lung carcinoma.

Methods: Herein we analyzed the antitumoral effect of usnic acid on the same cell line. We analyzed the gene expression results on *APOPT1*, *CYCS*, *APAF1*, *CASP3/9*, *TNF*, *BCL2*, *BCL2L1* and *AIFM1* genes which have possible role on cell apoptosis.

Results: Usnic acid has stimulatory effect on *APOPT1*, *CYCS*, *APAF1*, *CASP3*, *CASP9* gene expressions in our experiments.

Conclusion: Usnic acid has antitumoral effect on cancer cells, affecting on mitochondrial apoptotic genes.

Keywords : Usnic acid, cell growth, apoptosis, gene, antitumoral affect.

ÖZET

Amaç: Usnik asit likenlerde bulunan halen rolü tam olarak ortaya konamamış bir sekonder metabolittir. Usnik asit içeren liken ekstraktları tıpta, parfüm yapımında, kozmetikte ve ekolojide kullanılmaktadır. Usnik asit insan akciğer kanserinde hücre büyümesini durdurmakta, hücre döngüsünün durdurulması ve apoptozu aktive etmektedir. Çalışmamızda aynı hücre hattında usnik asitin antitümöral etkisi bakılmıştır.

Yöntem: Hücre apoptozunda rolü olan *APOPT1*, *CYCS*, *APAF1*, *CASP3/9*, *TNF*, *BCL2* ve *BCL2L1* genlerinin ekspresyon analizine bakıldı.

Bulgular: Usnik asitin *APOPT1*, *CYCS*, *APAF1*, *CASP3*, *CASP9* genleri üzerinde uyarıcı etkisi olduğu saptandı.

Sonuç: Veriler usnik asitin kanser hücreleri üzerinde mitokondrial apoptotik genler aracılığı ile antitümöral etki yaptığını ortaya koymaktadır.

Anahtar sözcükler: Usnik asit, hücre büyümesi, apoptoz, gene, antitümöral etki.

INTRODUCTION

Lichens are a world-widespread consortium of fungal and photosynthetic partners. Usnic acid is uniquely found in lichens, and is especially abundant in genera such as *Alectoria*, *Cladonia*, *Usnea*, *Lecanora*, *Ramalina* and *Evernia*. Usnic acid is one of the most common and abundant lichen metabolites, well known as an antibiotic, but also endowed with several other interesting properties¹. Many lichens and extracts containing usnic acid have been utilized for medicinal, perfumery, cosmetic as well as ecological applications. Usnic acid as a pure substance has been formulated in creams, toothpaste, mouthwash, deodorants and sunscreen products, in some cases as an active principle, in others as a preservative. In addition to antimicrobial activity against human and plant pathogens, usnic acid has been shown to exhibit antiviral, antiprotozoal, antiproliferative, anti-inflammatory and analgesic activity. Ecological effects, such as antigrowth, antiherbivore and anti-insect properties, have also been demonstrated². Usnic acid has antimicrobial, antiprotozoal, antiviral, antiproliferative, anti-inflammatory, analgesic and antipyretic activity³. Anti-inflammatory, analgesic and antipyretic effects have been suggested to be linked to inhibition of prostaglandin synthesis owing to the uncoupling effects on oxidative phosphorylation³. The toxicology, the in vitro effects and the mechanism of action of usnic acid need to be investigated in greater detail in order to reach clinical trials and to allow further applications¹. Usnic acid has antimitotic and antiproliferative activities against normal and malignant human cells⁴. Usnic acid (50 mg/ml) was shown to reduce cell counts of leukemic (K-562) and endometrial carcinoma cell lines (Ishikawa, HEC-50) when exposed to the cultures for 21 hours⁵. Usnic acid inhibits growth and induces cell cycle arrest and apoptosis in human lung carcinoma A549 cells⁶. Some studies have shown the anticancer potential of usnic acid; however, its efficacy and associated mechanisms are yet to be fully explored⁶.

Apoptogenic 1, Mitochondrial (APOPT1) gene is a protein that localizes to the mitochondria, where it stimulates the release of cytochrome c, thereby promoting programmed cell death⁷. *Cytochrome C, Somatic (CYCS)* gene encodes a small heme protein that functions as a central component of the electron transport chain in mitochondria. The encoded protein associates with the inner membrane of the mitochondrion where it accepts

electrons from cytochrome b and transfers them to the cytochrome oxidase complex. This protein is also involved in initiation of apoptosis⁸. *Apoptotic Peptidase Activating Factor 1 (APAF1)* gene encodes a cytoplasmic protein that initiates apoptosis⁹. *Caspase 3 (CASP3)* and *Caspase 9 (CASP9)* genes encode proteins which are the members of the caspase family. Sequential activation of caspases plays a central role in the execution-phase of cell apoptosis¹⁰. Singt et al. reported that usnic acid inhibits cell growth involving G0/G1 phase cell cycle arrest and induces cell death via mitochondrial membrane depolarization and induction of apoptosis in A549 cells⁶. Tumor Necrosis Factor (TNF) protein as a cytokine is involved in the regulation of a wide spectrum of biological processes including cell proliferation, differentiation, apoptosis, lipid metabolism, and coagulation¹¹. *BCL2, Apoptosis Regulator (BCL2)* gene encodes an integral outer mitochondrial membrane proteins that block the apoptotic death¹². *BCL2 Like 1 (BCL2L1)* gene is located at the outer mitochondrial membrane, and have been shown to regulate outer mitochondrial membrane channel (VDAC) opening¹³. *Apoptosis Inducing Factor, Mitochondria Associated 1 (AIFM1)* gene encodes a flavoprotein essential for nuclear disassembly in apoptotic cells¹⁴. So, we decided to find out the alterations of *APOPT1*, *CYCS*, *APAF1*, *CASP3/9*, *TNF*, *BCL2*, *BCL2L1* and *AIFM1* genes.

Herein, we assessed the anticancer potency of usnic acid. We tried to find out the molecular gene alterations on the *APOPT1*, *CYCS*, *APAF1*, *CASP3/9*, *TNF*, *BCL2*, *BCL2L1* and *AIFM1* genes in usnic acid treatment on A549 cells. Usnic acid treatment decreased total cell number and enhanced cell death in tumor cell culture. Usnic acid treatment increased the expressions of *APOPT1*, *CYCS*, *APAF1*, *CASP3*, *CASP9* genes. The proliferation of tumor cells was decreased tumor cell's growth and proliferation by using mitochondrial apoptotic genes.

MATERIAL AND METHODS

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Preparation of Extract Usnic Acid Solutions

We supplied usnic acid from Sigma Aldrich [329967-98% (Aldrich)]. We used Physiological Saline Solution (PSS) for dilutions in control and

study groups. The usnic acid solutions were prepared in different concentrations (1 µg/ml, 10 µg/ml, 50 µg/ml, 100 µg/ml, 200 µg/ml, 500 µg/ml).

Cell Culture of Human lung carcinoma A549

A549 cells was used for finding the apoptotic mechanisms in usnic acid at gene level. A549 cells (ATCC No: CCL-185) were cultured in flasks in RPMI 1640 (Sigma-Aldrich-R8758) containing 10% fetal bovine serum (FBS) (BiochromAG, Germany), 2% penicillin and streptomycin (Biological Industries, Israel). The cells were maintained in RPMI 1640 plus 10% FBS for at least 1 week before using them for experiments. Cells were maintained at 37 °C in a humidified atmosphere of 95% air–5% CO₂ in Heraeus incubator (Hanau, Germany). Cell cultures of cell line were applied due to classical standards (15). In control group, 50µl PSS was applied in each culture flasks. In studying group, 50 µl (1 µg/ml, 10 µg/ml, 50 µg/ml) usnic acid solution was added in each culture flasks. The total RNAs were in the first 24th hour by using RNA isolation kit (Roche RNA isolation kit).

Cytotoxicity Assay

Usnic acid solution in each concentration (1 µg/ml, 10 µg/ml, 50 µg/ml, 100 µg/ml, 200 µg/ml, 500 µg/ml) was added as 50µl in a culture flask. Toxic dosages of all two cell culture groups were determined by using “MTT (2,3-bis(2-methoxy-4-nitro-5-sulfophenyl)-5-[(phenylamino)carbonyl]-2H-tetrazolium hydroxide) assay” (16). LD50 value of usnic acid was found as 50 µg/ml in A549 cells. Due to MTT assay results, three different usnic acid concentrations (LD50 value of usnic acid and two lower dosages of usnic acid-1 µg/ml, 10 µg/ml, 50 µg/ml) were chosen and studied in our experiments.

Cell viability assay in cell cultures

Trypan blue exclusion test was applied to assess cell viability with trypan blue solution. The cell viability was examined under an inverted microscope (40X magnification). The cells were identified as degenerated (blue-stained) and survived (unstained) (Sigma-Aldrich T8154 Trypan Blue solution)¹⁶. In all well plates, the surviving ratios of the cells were found over 88%.

RNA Isolation and C-DNA synthesis from cell cultures

Total RNA isolation was performed from each sample separately (in three times). Cells were harvested from flasks by trypsinization and collected in a tube. RNAs were converted to c-

DNAs by using the c-DNA synthesis kit (RevertAid cDNA Synthesis Kit, Fermentas). c-DNA samples quality controlled by %2'lik agarose gel under ultraviolet light.

Reverse Transkriptaz Polimeraz Chain Reaction (RT-PCR); The gene expressions of all selected genes were studied in this panel. The studied genes were *β Actin (ACTB)* (OMIM: 102630) (for control), *APOPT1-Apoptogenic 1, Mitochondrial* (OMIM: 616003), *CYCS-Cytochrome C, Somatic* (OMIM: 123970), *APAF1 Gene-Apoptotic Peptidase Activating Factor 1* (OMIM: 602233), *CASP3-Caspase 3* (OMIM: 600636), *CASP9-Caspase 9* (OMIM: 602234), *TNF- Tumor Necrosis Factor* (OMIM: 191160), *BCL2-BCL2 Apoptosis Regulator* (OMIM: 151430), *BCL2L1-BCL2 Like 1* (OMIM: 600039), *AIFM1-Apoptosis Inducing Factor, Mitochondria Associated 1* (OMIM: 300169). All c-DNA's were used in RT-PCR. All primer information used in PCR are taken from <http://pga.mhg.harvard.edu/primerbank/> site. Each RT-PCR reaction was performed in 20 µl [10 µl 2 X SYBR, 5 µl c DNA, 1 µl primer, 3 µl d H₂O] (Roche Applied Science: LightCycler® 480 System). SYBR Green PCR master mix was obtained from Applied Biosystems. RT-PCRs were applied for 6 times for each gene separately. The gene expression levels of all genes in A549 cells in the first 24th hour were found. The gene expression results shown on the table were found due to control group (*β Actin*) results. The results on the table were found by reducing the control results in each condition. Mean values were obtained in all groups. The critical quantification cycle (Cq) value was defined based on the number of cycles at which the fluorescence signal can be detected above a threshold value and inversely correlated to the initial amount of DNA present in the PCR reaction¹⁷.

Statistical Analysis; Student's t test (one sample t test) was used for two-group comparisons by using SPSS programme. The tests results (p values) represent two group comparisons among the control and the usnic acid studied groups (Table 1).

RESULTS

In order to test the anti-tumour properties of usnic acid on human cells, three different concentrations were used on A549 cells. In our experiments, usnic acid in concentrations 10 µg/ml and 50 µg/ml inhibited the proliferation of tumoral cells. Usnic acid in 1 µg/ml concentration had no effect on tumoral cells. The results in 10 µg/ml and 50 µg/ml concentrations indicated that usnic acid strongly inhibits tumoral cell growth and proliferation in

certain concentrations. In cell viability assay results, cell viability of usnic acid applied in A549 cell culture group were found lower (88%-10 µg/ml usnic acid and 90%-50 µg/ml usnic acid) than control group (%91).

In our study, the gene expression differentiations were detected by Real-time PCR. The results were shown on table. Due to expression values of selected genes which participate on apoptotic pathways, no change was occurred in uses of 1 µg/ml usnic acid concentration. In the analyses of 1 µg/ml usnic acid concentration, no statistical difference was observed due to control results ($p \geq 0,05$), except in CASP3 gene analyses results

(Table). The *APOPT1*, *CYCS*, *APAF1*, *CASP3* and *CASP9* genes had higher gene expression results in usnic acid 10 µg/ml and 50 µg/ml concentrations than control. In *TNF*, *BCL2*, *BCL2L1* and *AIFM1* gene analyses, no significant difference was observed in RT-PCR analyses ($p \geq 0,05$) (Table).

So, antiproliferative effect of usnic acid on human lung cancer tumoral cells in our experiments was found in our analyses. Usnic acid in certain concentrations did antitumoral effect on these cell line using *APOPT1*, *CYCS*, *APAF1*, *CASP3* and *CASP9* genes which have role on tumor apoptotic pathways.

Table 1. The gene expression results of usnic acid in selected genes participate on apoptotic pathways in our experiment.

Genes	Control	Usnic acid (1 µg/ml)	p value	Usnic acid (10 µg/ml)	p value	Usnic acid (50 µg/ml)	p value
<i>APOPT1</i>	0,096±0.03	0,090±0.04	$p \geq 0,05$	0,245±0.09	$p < 0,05$	0,986±0.22	$p < 0,05$
<i>CYCS</i>	0,004±0.07	0,006±0.12	$p \geq 0,05$	0,011±0.12	$p < 0,05$	0,009±0.02	$p < 0,05$
<i>APAF1</i>	1.112±0.03	1.122±0.11	$p \geq 0,05$	2.132±0.16	$p < 0,05$	2.144±0.33	$p < 0,05$
<i>CASP3</i>	0.122±0.17	0.142±0.10	$p < 0,05$	0.152±0.09	$p < 0,05$	0.202±0.09	$p < 0,05$
<i>CASP9</i>	1.899±0.27	1.802±0.24	$p \geq 0,05$	2.802±0.44	$p < 0,05$	3.002±0.64	$p < 0,05$
<i>TNF</i>	0.060±0.03	0,060±0.01	$p \geq 0,05$	0.050±0.01	$p \geq 0,05$	0.055±0.02	$p \geq 0,05$
<i>BCL2</i>	0,099±0.09	0,101±0.10	$p \geq 0,05$	0,095±0.13	$p < 0,05$	0,098±0.13	$p \geq 0,05$
<i>BCL2L1</i>	0,637±0.19	0,629±0.20	$p \geq 0,05$	0,631±0.08	$p \geq 0,05$	0,632±0.09	$p \geq 0,05$
<i>AIFM1</i>	0,009±0.07	0,008±0.03	$p \geq 0,05$	0,008±0.02	$p \geq 0,05$	0,009±0.02	$p \geq 0,05$

DISCUSSION

One of the ways of searching potentially new anti-cancer drugs is testing the various naturally synthesized compounds like lichens to be effective against various cancer in vitro models. Usnic acid as a lichen metabolite is known to exert antimetabolic and antiproliferative activities against normal and malignant human cells. Usnic acid blocks cell cycle progression and induces cell death through apoptosis¹⁸. Usnic acid treatment of breast adenocarcinoma MCF-7 and human non-small cell carcinoma H1299 cells did not result in any morphological changes in microtubules or increase in the mitotic index. O'Neill et al. explained that the antineoplastic activity of usnic acid was not related to alterations in the formation and/or stabilisation of microtubules¹⁹. In comparison with parietin and gyrophoric acid, the suppression of viability and cell proliferation by usnic acid or atranorin was found to be more efficient at equitoxic doses. Usnic acid and atranorin correlated more strongly with an increased number of floating cells or a higher apoptotic index. Bačkorová et al. found the cell cycle distribution is important for the drug sensitivity¹⁸. Like observed in literature, usnic acid had antiproliferative effect on human tumor cells in

our analyses. We observed the antitumoral effect on certain usnic acid dosages.

Apoptosis is another mechanism affected on usnic acid's antitumoral effect. Brisdelli et al.'s study was pointed out this effect of usnic acid. In this study, the effects of six lichen metabolites (diffractaic acid, lobaric acid, usnic acid, vicanicin, variolaric acid, protolichesterinic acid) were analyzed on proliferation, viability and reactive oxygen species level towards three human cancer cell lines [MCF-7, HeLa (cervix adenocarcinoma) and HCT-116 (colon carcinoma)]. Their study revealed that the antiproliferative activity of protolichesterinic acid in HeLa cells. It was related to its ability to induce programmed cell death involving caspases²⁰. Apoptosis is caused by proteases, known as "caspases," which specifically target cysteine aspartyl. These caspases activate themselves and each other. Within these proteolytic cascades, caspases can be positioned as either upstream "initiators" or downstream "effectors" of apoptosis. Several pathways for activating caspases exist²¹. First, there are thirty members of the TNF family receptors; eight contain a so-called death domain in their cytosolic tail²². Second is the intrinsic pathway, in which

mitochondria induces apoptosis by releasing cytochrome c into the cytosol. The released cytochrome c assembles a multiprotein caspase-activating complex, referred to as the “apoptosome”²³. A third pathway for apoptosis induction is specific to natural killer-NK cells, which spray apoptosis-inducing protease, granzyme B (GraB), onto target cells²⁴. In our analyses, *APOPT1*, *CYCS*, *APAF1*, *CASP3* and *CASP9* genes had higher gene expression results than control. So usnic acid affect on apoptotic genes in it's antitumoral effect (Table).

APOPT1 gene encodes a protein that localizes to the mitochondria, where it stimulates the release of cytochrome c, thereby promoting programmed cell death²⁵. *CYCS* gene encodes a small heme protein that functions as a central component of the electron transport chain in mitochondria. This protein is also involved in initiation of apoptosis²⁶. *APAF1* gene encodes a cytoplasmic protein that initiates apoptosis. This protein contains several copies of the WD-40 domain, a caspase recruitment domain (CARD), and an ATPase domain (NB-ARC). Upon binding cytochrome c and dATP, this protein forms an oligomeric apoptosome. The apoptosome binds and cleaves caspase 9 preproprotein, releasing its mature, activated form. Activated caspase 9 stimulates the subsequent caspase cascade that commits the cell to apoptosis^{26, 27}. *Caspase 3* gene encodes a protein which is a member of the cysteine-aspartic acid protease (caspase) family. Sequential activation of caspases plays a central role in the execution-phase of cell apoptosis²⁷. *Caspase 9* gene encodes a member of the cysteine-aspartic acid protease (caspase) family. Sequential activation of caspases plays a central role in the execution-phase of cell apoptosis. Caspases exist as inactive proenzymes which undergo proteolytic processing at conserved aspartic residues to produce two subunits, large and small, that dimerize to form the active enzyme. This protein can undergo autoproteolytic processing and activation by the apoptosome, a protein complex of cytochrome c and the apoptotic peptidase activating factor 1; this step is thought to be one of the earliest in the caspase activation cascade²⁷. The gene expression analysis of *APOPT1*, *CYCS*, *APAF1*, *Caspase 3* and *9* genes were found as high in our experiments which represent the role of usnic acid in apoptosis.

TNF gene encodes a multifunctional proinflammatory cytokine that belongs to the tumor necrosis factor (TNF) superfamily. This cytokine is mainly secreted by macrophages. This cytokine is involved in the regulation of a wide spectrum of biological processes including cell

proliferation, differentiation, apoptosis, lipid metabolism, and coagulation²⁸. *BCL2* encodes an integral outer mitochondrial membrane protein that blocks the apoptotic death of some cells such as lymphocytes. Constitutive expression of *BCL2*, such as in the case of translocation of *BCL2* to Ig heavy chain locus, is thought to be the cause of follicular lymphoma²⁹. *BCL2L1* encoded by this gene belongs to the BCL-2 protein family. BCL-2 family members form hetero- or homodimers and act as anti- or pro-apoptotic regulators that are involved in a wide variety of cellular activities. The proteins encoded by this gene are located at the outer mitochondrial membrane, and have been shown to regulate outer mitochondrial membrane channel opening which is important in apoptosis of cell^{28, 29}. *AIFM1* gene encodes a flavoprotein essential for nuclear disassembly in apoptotic cells, and it is found in the mitochondrial intermembrane space in healthy cells. Induction of apoptosis results in the translocation of this protein to the nucleus where it affects chromosome condensation and fragmentation. In addition, this gene product induces mitochondria to release the apoptogenic proteins cytochrome c and caspase 9²⁹. In our experiment, no increase was observed in the expression levels of *TNF*, *BCL2*, *BCL2L1* and *AIFM1* genes by using usnic acid. All of these results support the apoptotic role of usnic acid.

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