

Review - Derleme

Ischemic preconditioning and postconditioning in cardiovascular surgery

Kardiyovasküler cerrahide iskemik önkoşullanma ve sonradan koşullanma

İlker Şengül, Demet Şengül

Department of General Surgery (Assist Prof. İ. Şengül, MD) Giresun University School of Medicine ,TR-28100 Giresun, Department of Pathology (Specialist D. Şengül, MD) Prof. Dr. A. İlhan Özdemir State Hospital TR-28100 Giresun.

Abstract

Despite the technical improvements in the procedures of cardiovascular surgery; significant morbidity and mortality still continue in the patients undergoing cardiovascular surgery, especially in the high risk ones. Models of “conditioning” are suggested and evaluated as preserving the heart itself against ischemia - reperfusion injury which is an important cause of myocardial injury created by the mentioned surgery and these models have been used in lots of experimental / clinical studies. In the mechanism of ischemic preconditioning; various systemic, chemical, physical stimuli, the substances affecting on the paired receptors of G - protein, proinflammatory substances, changes in the oxygen tension and various hypotheses were suggested such as protein kinase activation and activation of myocardial potassium-ATP (KATP) channels. Lots of affirmative cardiovascular interventions such as coronary artery bypass surgery, cardiac valve replacement, transplantation in vivo and vitro have been reported to benefit from the myocardial preservation effect of ischemic preconditioning. Postconditioning have been widely used in various experimental / clinical studies such as acute myocardial infarction, ventricular defibrillation, repair of tetralogy of Fallot and transplantation. Despite the fact that these “conditioning” phenomenons are promising, the demand for larger, multicenter, randomised, placebo - controlled experimental / clinical studies in this subject is increasing day by day.

Key words: Cardiovascular surgical procedure; conditioning; ischemic preconditioning; postconditioning; ischemia; reperfusion.

Özet

Kardiyovasküler cerrahi prosedürlerinde teknik ilerlemelere rağmen, hala kardiyovasküler cerrahiye giden - özellikle yüksek riskli – hastalarda, belirgin morbidite ve mortalite süregelmektedir. Anılan cerrahinin yarattığı miyokard hasarının ileri gelen nedeni olan iskemi – reperfüzyon hasarına karşı kalbin kendini koruması olarak bakılan “koşullanma” modelleri öne sürülmüş ve deneysel / klinik birçok çalışmada kullanılmıştır. İskemik önkoşullanmanın mekanizmasında; çok miktarda sistemik, kimyasal, fiziksel uyarın, G - proteinin çifteleşmiş reseptörleri üzerine etki eden maddeler, proinflamatuvlar maddeler, oksijen tansiyonundaki değişimler ile birlikte protein kinaz aktivasyonu ve miyokardiyal potasyum - ATP (KATP) kanal aktivasyonu gibi daha birçok hipotez ileri sürülmüştür. İskemik önkoşullanmanın miyokard koruyuculuğu eylemi ile ilgili in vivo ve in vitro olmak üzere koroner arter “bypass” cerrahisi, kalp kapak replasmanı, transplantasyon gibi birçok olumlu kardiyovasküler cerrahi girişim raporlanmıştır. Sonradan koşullanma ise akut miyokard infarktüsü, ventriküler defibrilasyon, Fallot tetralojisi onarımı ve transplantasyon gibi alanlardaki deneysel / klinik birçok çalışmada geniş kullanım alanı bulmuştur. Bununla birlikte; bu “koşullanma” fenomenleri umut verici olsa da daha geniş, çok merkezli, randomize, placebo - kontrollü, deneysel ve klinik çalışmalara gereksinim gün geçtikçe artmaktadır.

Anahtar sözcükler: Kardiyovasküler cerrahi prosedür; koşullanma; iskemik önköşullanma; sonradan koşullanma; iskemi; reperfüzyon.

Geliş tarihi/Received: 27 Nisan 2010; **Kabul tarihi/Accepted:** 27 Mayıs 2010

İletişim Adresi:

Dr. İlker Şengül Genel Cerrahi Anabilim Dalı Giresun Üniversitesi Tıp Fakültesi TR-28100
Giresun E-posta: dr.ilker52@mynet.com

Introduction

It has been observed that the patients undergoing cardiovascular surgery are still at great risk despite the various technical improvements and the results of cardiovascular (CV) surgical procedures beginning from the 1950s and 1960s. CV arrest is one of the critical parts of the CV surgery (CVS) and at the same time it means CV ischemia.

During the 1970s, in cardioplegic arrest; the heart was arrested by infusing a cold solution (4-8°C) into coronary circulation containing high concentrations of potassium which acts like an arrest forming agent. This method has been made a part of the CV repertoire since then.

Initially the solution was crystalloid based which later was replaced by blood based cardioplegia. Today the most commonly used form is the blood based one which has been proven in many studies to be superior to crystalline based ones.

Blood based CP can be applied as cold warm or hot but hot CP is the most popularly used.

Advanced regimens are still needed for myocardial protection of patients who are old, have accompanying diseases and who have significantly depressed CV function in order that they can undergo CVS. Ischemic preconditioning (IPC) and postconditioning (Postcon) a latest determined phenomenon can increase myocardial protection during cardiac surgery. The adaptive effect of them on myocardium was observed to be decreasing infarct area and functional improvement [1].

Ischemic preconditioning in cardiovascular surgery

Various systemic, chemical [2] and physical [3] stimuli have been suggested in the mechanism of IPC together with opioid and adenosine which act on the coupled receptors of G protein; proinflammatory substances such as lipopolysaccharides, TNF α and monophosphoryl lipid A and changes in oxygen tension in the form of anoxia, hypoxia and hyperoxia. A large variety of drugs and anesthetics used routinely in clinical practice form IPC effect. Various hypotheses have been also suggested such as protein kinase activation and myocardial potassium-ATP (K_{ATP}) channel activation [4-17].

There are some clinical studies in which IPC is used in open heart surgery. However the surgeons remained reluctant for classical IPC. They do not wish repeatedly clamping of aorta because of the risk of atheroembolism from calcifications in ascending aorta [18]. Additionally, IPC can itself become harmful by prolonging the surgical procedure for extra 15-30 minutes. Conflicting results were reported for the cardio protective effect of IPC when it was combined with hypothermia and/or CP [19, 20]. The brittle side of this matter is that IPC can be in relation with estradiol and does not clarify the answer of the question "Can it be useful for the female population?" [21-24]. All considerations are valid for Postcon except prolonging the duration of the procedure [1].

It has been observed that the healthiness and youngness of animals in laboratory animal

studies concerning IPC and Postcon and the presence of accompanying diseases such as coronary heart disease, diabetes, heart failure and the age of the patient in clinical human studies can affect the results. The metabolic changes induced by hypercholesterolemia and atherosclerosis were investigated whether they can complicate IPC by causing chronic ischemic heart disease [16]. Szers et al [25] have not observed myocardial protection in rats fed on high cholesterol diet for 24 weeks, while Li et al [26] have demonstrated that IPC provides more protection in advanced age rats having overt atherosclerosis due to apolipoprotein E/LDL gene defect when compared with young rats with normal vascular structure.

Additionally Valen et al [27] have shown that rats with serious atherosclerosis experiencing in vivo spontaneous infarction were protected from ex vivo global ischemia. The adaptation way to this kind of ischemia was thought to be a phenomenon formed by natural ways and this condition is met in atherosclerosis types analogous to unstable angina. For this reason, atherosclerosis is not seen as a spontaneous factor that prevents IPC. Unstable coronary syndromes are progressed by systemic inflammatory events [28] or accompanied by these events and can mimic IPC like a clinical analogue. It has been shown in various studies that the outcome is better in patients with unstable angina [UA] than in patients experiencing acute myocardial infarction without angina; the mortality, the incidence of shock and serious congestive heart failure is also lower, the infarct areas are smaller in correlation with the decrease in creatinine phosphokinase activity and lesser Q wave activity were also observed in patients with unstable angina [29, 30]. In Tampere Finland; sensitive parameters for IPC concept were evaluated and “infarct area” was defined as “gold standard” [1].

The molecular changes observed in patients who have UA in CV tissue and undergo open cardiovascular surgery are the same as the changes in animals with experimentally induced IP [31]. Postoperative functional myocardial protection was observed in UA patients but an additional IP was reported not to bring any benefit [32]. It is very difficult to discriminate the reason for the adaptation to ischemia whether it is due to intermittent hypoxia and reoxygenation episodes or due to intravenous glycerin during treatment [33, 34]. Whatever the mechanism is patients who experience UA or myocardial infarction preoperatively, may not benefit from IPC or Postcon [1].

The initial studies about IPC in open heart surgery was performed by Yellon et al in 1993 by performing intermittent clamping for CV protection and showing protected CV ATP levels in patients undergoing coronary artery “bypass” surgery (CABS). Later this same group and one year after Szamagala et al [36] have shown decreased CV troponin T levels in this model [37].

It has been discussed that cardioplegic bypass during CABS cannot bring any benefit other than self-induced IPC [38] and Burns et al [39] have shown that IPC is induced by cardioplegic bypass via stimulating adrenergic and adenosin-1 receptors. Additionally like sevofluran which is known have an IPC effect, it activates kinase cascades which is related with the opening up of potassium channels [38]. Cardioplegic bypass can also induce general inflammatory response including cytokines like TNF α (tumor necrosis factor-alpha) and ROS (reactive oxygen species) production [40] and this can trigger the response of IP [41].

Illes and Swoyer [42] have found increased CV function and decreased inotropic support after IPC. Also in other studies IPC has been reported to be protective in patients undergoing valve surgery and coronary artery bypass graft surgery [43, 44].

IPC was shown to provide more cardiac protection when compared with the pharmacological preconditioning induced by adenosine A-1 receptors [45]. Finland Tampere cardiac surgery group has investigated prospectively ischemic preconditioning in a series of randomized patients undergoing coronary artery bypass surgery and found data yielding that IPC increases the functions of both two left and right ventricles in the

early postoperative period [46, 47]. It has been observed that the IPC response in the elderly patients (Age: >68) is not as clear as the response in younger (Age: <68) patients [48]. Wu et al reported that UA induced IPC during the last 48 hours before surgery increases the postoperative hemodynamic of patients [32]. IP was shown to significantly decrease postoperative ventricular tachycardia and ventricular fibrillation [49] and incidence of postoperative atrial fibrillation in patients with three vessel disease after CABS [50]. The suppression of ventricular tachycardia during early reperfusion and 24 hours after surgery suggests that both the early and late effects of IPC are observed in patients undergoing IPC surgical protocol. In another study performed by Wu et al [51] cardioplegia induced apoptosis was not inhibited by IPC. Diazoxide induced myocardial protection during CABS suggests that ATP-dependent potassium channels are important in IPC during CVS [52].

Investigators from Finland have clamped aorta in its whole horizontal axis for 2 minutes before intermittent cold blood based cardioplegia and left to reperfusion for 3 minutes and repeated this for one time and showed a protection consistent with this regimen [44, 49]. Recently Venugopal et al. [53] have demonstrated in a randomized controlled study of IPC model in the arm. They reported that myocardial destruction was reduced in patients undergoing CABS by using the IPC model and that non-invasive cardio protective technique had a wide use of area clinically. However various anesthetics, different patient populations, the abundance of cardioplegic solutions become factors that make IPC complicated in CVS [1].

Postconditioning in cardiovascular surgery

In the recent years the phenomenon of Postcon defined as short term ischemic attacks observed after long term ischemia has been found to decrease infarct area as effective as IPC in a dog model [54] but has a lesser effect than IPC in a rat model [55]. Another aspect of ischemia-reperfusion injury is the reperfusion arrhythmia [56]. In a recent study, Zhang et al [57] have shown that Postcon of sevoflurane can convert persistent ventricular fibrillation to normal rhythm.

Postcon has initially been studied in the heart [54] followed by the liver [58] and recently in different organs such as the brain [59-61], kidney [62-64], the straightened muscle[65], skin flap [66] and in small intestines [67-71]. We investigated the effect of Postcon on intestinal ischemia reperfusion (I/R) injury (IRI) via inhibition of the events in the first few minutes in 5 group and 3 separate Postcon models (PC - 3, PC - 6 and delayed PC [DPC]) of rats. After applying 30 minutes of global ischemia to the superior mesenteric artery to each rat after laparotomy, 3 cycles (PC - 3) and 6 cycles (PC-6) lasting 10 minutes each and 3 cycles after one minute (DPC) were performed in the reperfusion period to maintain intermittent ischemia, after then we left them to global reperfusion for period of 120minutes. By using Postcon models, we demonstrated that infarct area (serum total creatine kinase [CK]), lipid peroxidation (tissue malonyl dialdehyde [MDA]) and the destruction in ileum morphology (Histopathological score) have decreased [67]. Postcon has been shown to decrease infarct area of a myocardial infarct model in dogs, rats, [72-74] and rabbits [75] and also decrease reperfusion arrhythmias of isolated rat heart [57]. After hypoxia-reoxygenation, alive cell count of neonatal rat cardiomyocytes were found to be increased by hypoxic PC [76].

The initial clinical studies about Postcon were conducted by Laskey [77] and Staat et al [78] in patients with acute myocardial infarction. In these studies it was reported that low pressure application induced in the angioplasty balloon for 4 times each lasting 1 minutes increases myocardial infarction, decreases myocardial infarction area both in the acute and during the 6 months period and increases the left ventricular ejection fraction during the 1 year period [77-79]. In 2007, Luo et al. [80] first conducted a randomized controlled study in which PC was investigated in surgery patients. This study included 24 children

with Fallot tetrology and it was found that the PC group required significantly lesser inotrop during the first 24 hours and 50% lower levels of troponin I ($p = 0,05$) and 34% lower levels of CK-MB ($p = 0,034$) were detected. In 2008 the same group reported similar favorable results regarding the adult valve replacement surgery [81]. Likewise in 2009; Li et al [82] suggested in their study including 99 patients who underwent Fallot tetrology repair that Postcon has beneficial effects on morbidity, ventilation time, hospitalization period in intensive care unit, troponin I and lactate release and inotrop requirement. Additionally the myocardial protective effect of Postcon in children who underwent cardioplegic arrest has first been demonstrated by the same group [83].

Cardiac transplantation surgeons can use Postcon. The preservation period of cardiac allograft can often exceed the protective effects of early IPC. For this reason Postcon can play an important role in allograft protection following storage and transplantation. Additionally, Postcon was shown to induce a high degree cardio protective effect in a long, cold (4 hours / 4 °C) ischemia protocol and also it was demonstrated that Postcon induces low oxygen production formed by a partial decrease in NADPH oxidase activity [84, 85].

Conclusion

There is still significant morbidity and mortality in patients undergoing CVS especially in high risk patient groups despite the recent developments in myocardial protective techniques and evolution of off-pump CABS. The major cause of myocardial injury during CVS is acute ischemia reperfusion injury (71) Conditioning which is regarded as self-protection of heart from this injury can be divided as IPC which means a cardioprotective intervention before myocardial ischemia and Postcon which is applied after ischemia. Until today various experimental studies have been carried out about this two phenomenon. Besides, although there are clinical, surgical and transplantation related articles, most of the ischemia is often not predictable. So the benefit of IPC is limited and Postcon seems to be more promising. For this reason, as the clinical application areas of Postcon are considered especially when the large patient group with myocardial infarction is taken account of Postcon becomes more attractive than IPC. Additionally, both in vivo and in vitro studies in the literature have demonstrated favorable results about IPC in various CV interventions such as CABS, cardiac valve replacement and transplantation and have found a large area of application.

In conclusion, we can say that although these conditioning phenomena against IRI seems to be promising, there is still a need of large, randomized, placebo-controlled multicenter studies both experimental and clinical.

References

1. Valen G, Vaage J. Pre- and postconditioning during cardiac surgery. *Basic Res Cardiol* 2004-5;100:179-86.
2. Domenech R, Macho P. Non- ischemic myocardial preconditioning. *Mol Cell Biochem* 1998;186:201-3.
3. Yellon MD, Downey JM. Preconditioning the myocardium: from cellular physiology to clinical cardiology. *Physiol Rev* 2003;83:1113-51.
4. Sivaraman V, Hausenloy DJ, Kolvekar S, Hayward M, Yap J, Lawrence D, Di Salvo C, Yellon DM. The divergent roles of protein kinase C epsilon and delta in simulated ischaemia-reperfusion injury in human myocardium. *J Mol Cell Cardiol* 2009; 46: 758-64.
5. Baxter GF, Ferdinand P. Delayed preconditioning of myocardium: current perspectives. *Basic Res Cardiol* 2001; 96: 329-44.
6. Budas GR, Jovanovic S, Crawford RM, Jovanovic A. Hypoxia induced preconditioning in adult simulated cardiomyocytes is mediated by opening and trafficking of sarcolemmal K-ATP channel. *FASEB J* 2004;18: 1046-8.
7. Laude K, Beauchamp P, Thuillez C, Richard V. Endothelial effect of preconditioning. *Cardiovasc Res* 2002; 55: 466-73.
8. Marber MS. Ischemic preconditioning in isolated cells. *Circ Res* 1986;86:926-31.

9. Murphy E. Primary and secondary signaling pathways in early preconditioning converge in mitochondria to produce cardioprotection. *Circ Res* 2004; 94: 7-16.
10. Gao X, Zhang H, Takahashi T, Hsieh J, Liao J, Steinberg GK, Zhao H. The Akt signaling pathway contributes to postconditioning's protection against stroke; the protection is associated with the MAPK and PKC pathways. *J Neurochem* 2008; 105: 943-55.
11. Rehni AK, Singh N. Role of phosphoinositide 3-kinase in ischemic postconditioning-induced attenuation of cerebral ischemia-evoked behavioral deficits in mice. *Pharmacol Rep* 2007; 59: 192-8.
12. Wojtovich AP, Brookes PS. The complex II inhibitor atpentin A5 protects against cardiac ischemia-reperfusion injury via activation of mitochondrial KATP channels. *Basic Res Cardiol* 2009; 104: 121-9.
13. Lamendola P, Di Monaco A, Barone L, Pisanello C, Lanza GA, Crea F. [Mechanisms of myocardial cell protection from ischemia/reperfusion injury and potential clinical implications]. *G Ital Cardiol (Rome)* 2009; 10: 28-36. [Article in Italian]
14. Valen G. Cellular signaling mechanisms in adaptation of ischemia induced myocardial damage. *Ann Med* 2003; 35: 1-8.
15. Yellon DM, Dana A. The preconditioning phenomenon. A tool for scientist or a clinical reality? *Circ Res* 2000; 87: 543-50.
16. de Zeeuw S, Van den Doel MA, Duncker DJ, Verdouw PD. New insights into cardioprotection by ischemic preconditioning and other forms of stress. *Ann N Y Acad Sci*. 1999; 874: 178-91.
17. Şengül İ, Şengül D. İskemik ön koşullanma ve sonradan koşullanma mekanizmaları olarak intraselüler sinyalizasyon ve adenozin. *Cumhuriyet Tıp Derg* 2010; 32: 127-131.
18. Bopassa JC, Ferrera R, Gateau-Roesch O, Couture -Lepetit E, Ovize M. PI3-kinase regulates the mitochondrial transition pore in controlled reperfusion and postconditioning. *Cardiovasc Res* 2006; 69: 178-85.
19. Kolicassides KD, Galinanes M, Hearse D. Ischemic preconditioning, cardioplegia or both? *J Mol Cell Cardiol* 1994; 26: 1411-14.
20. Takeshima S, Vaage J, Löwbeer C, Valen G. Does hypothermia or hypokalemia influence the preconditioning response? *Scand Cardiovasc J* 1999; 33: 79-87.
21. von Arnim CA, Ettrich SM, Trümmel M, Riepe MV. Gender dependent hypoxic tolerance mediated via gender specific mechanisms. *J Neurosci Res* 2002; 68: 84-8.
22. Lee TM, Su SF, Tsai CC, Lee YT, Tsai CH. Cardioprotective effects of 17-β estradiol produced by activation of mitochondrial ATP-sensitive K channels in canine hearts. *J Mol Cell Cardiol* 2000; 32: 1147-58.
23. Shinmura K, Nagai M, Tamaki K, Bolli R. Gender and aging do not impare opioid-related preconditioning in rats. *Basic Res Cardiol* 2004; 99: 46-55.
24. Song X, Li g, Vaage J, Valen G. Effects of sex, gonadectomy and estrogen substitution on ischemic preconditioning and ischemia- reperfusion injury in mice. *Acta Physiol Scand* 2003; 177: 459-66.
25. Szekers L, Szilvassy Z, Ferdinand P, Nagy I, Karcsu S, Csáti S. Delayed cardiac protection against harmful consequences of stress can be induced in experimental atherosclerosis in rabbits. *J Mol Cell Cardiol* 1997; 29: 1977-83.
26. Li G, Tokuno S, Tahepol P, Vaage J, Löwbeer C, Valen G. Preconditioning protects the severely atherosclerotic mouse heart. *Ann Thorac Surg* 2001; 71: 1296-303.
27. Tokuna S, Hinokiyama K, Tokuna K, Löwbeer C, Valen G. Spontaneous ischemic events in the heart and brain precondition the hearts of severely atherosclerotic mice. *Atheroscler Thromb Vasc Biol* 2002; 22: 995-1001.
28. Mehta JL, Saldeen TGP, Rand K. Interactive role of infection, inflammation and traditional risk factors in atherosclerosis and coronary artery disease. *Circ Res* 1998; 31: 1217-25.
29. Abete P, Cacciatore F, Della Morte D, Mazzella F, Testa G, Galizia G, De Santis D, Longobardi G, Ferrara N, Rengo F. Joint effect of physical activity and body mass index on mortality for acute myocardial infarction in the elderly: role of preinfarction angina as equivalent of ischemic preconditioning. *Eur J Cardiovasc Prev Rehabil* 2009; 16: 73-9.
30. Sezer M, Nisancı Y, Ummam B, Ummam S, Pusuroglu H, Filorinalı N, Ozsaruhan O, Erzengin F. Effect of preinfarction angina pectoris on microcirculation in patients with reperfused acute myocardial infarction. *Acta Cardiol* 2004; 59: 25-31.
31. Valen G, Hansson GK, Dumitrescu A, Vaage J. Unstable angina activates myocardial heart shock protein 72, endothelial nitric oxide synthase and transcription factors NFkB and AP-1. *Cardiovasc Res* 2000; 47: 49-56.

32. Wu Z-K, Pehkonen E, Laurikka J, Honkonen EL, Kaukinen S, Tarkka MR. Protective effects of unstable angina in coronary artery bypass surgery. *Scand Cardiovasc J* 2000; 34: 486-92.
33. Heusch G. Editorial comment: Nitroglycerin and delayed preconditioning in humans. Yet another mechanisms for an old drug? *Circulation* 2001; 103: 2876-8.
34. Leesar MA, Stoddard MF, Drawn B, Jasti VG, Masten R, Bolli R. Delayed preconditioning-mimetic action of nitroglycerin in patients undergoing coronary angioplasty. *Circulation* 2001; 103: 2935-41.
35. Yellon DM, Alkhulaifi A, Pugsley WB. Preconditioning the human myocardium. *Lancet* 1993; 342: 276-8.
36. Szmagala P, Morawski W, Krejca M, Gburek T, Bochenek A. Evaluation of perioperative myocardial tissue damage in ischemic preconditioning human heart during aorto-coronary bypass surgery. *J Cardiovasc Surg* 1998; 39: 791-5.
37. Jenkins DP, Pugsley WB, Alkhulaifi AM, Kemp M, Hooper J, Yellon DM. Ischemic preconditioning reduces troponin T in patients undergoing coronary artery bypass surgery. *Heart* 1997; 77: 314-8.
38. Pouzet B, Lecharny JB, Dehoux M, Paquin S, Kitakaze M, Mantz J, Menasche P. Is there a place for preconditioning during cardiac operations in humans? *Ann Thorac Surg* 2002; 73: 843-8.
39. Burns PG, Krukenkap IB, Calderone CA, Kirvaitis RJ, Gaudette GR, Bukhari EA, Levitsky S. Does cardiopulmonary bypass alone elicit myoprotective preconditioning? *Circulation* 1995; 92 (9suppl): II447-51.
40. Paparella D, Yau TM, Yuong E. Cardiopulmonary bypass induced inflammation: pathophysiology and treatment. An update. *Eur J Cardiothorac Surg* 2002; 21: 232-44.
41. Sbrana S, Parri MS, De Filippis R, Gianetti J, Clerico A. Monitoring of monocyte functional state after extracorporeal circulation: a flow cytometry study. *Cytometry B Clin Cytom* 2004; 58: 17-24.
42. Illes RW, Swoyer KD. Prospective, randomized clinical study of ischemic preconditioning as an adjunct to intermittent cold blood cardioplegia. *Ann Thorac Surg*. 1998; 65: 748-52; discussion 752-3.
43. Li G, Chen S, Lu E, Li Y. Ischemic preconditioning improves preservation with cold blood cardioplegia in valve replacement patients. *Eur J Cardiothorac Surg* 1999; 15: 653-7.
44. Hausenloy DJ, Mwamure PK, Venugopal V, Harris J, Barnard M, Grundy E, Ashley E, Vichare S, Di Salvo C, Kolvekar S, Hayward M, Keogh B, MacAllister RJ, Yellon DM. Effect of remote ischaemic preconditioning on myocardial injury in patients undergoing coronary artery bypass graft surgery: a randomised controlled trial. *Lancet* 2007; 370: 575-9.
45. Teoh LK, Grant LR, Hulf JA, Pugsley WB, Yellon DM. The effect of preconditioning (ischemic and pharmacological) on myocardium necrosis following coronary artery bypass surgery. *Cardiovasc Res* 2002; 53: 175-80.
46. Wu Z-K, Tarkka M, Pehkonen E, Kaukinen L, Honkonen El, Kaukinen S. Beneficial effects of ischemic preconditioning on right ventricular function after coronary artery bypass grafting. *Ann Thorac Surg* 2000; 70: 1551-7.
47. Wu Z-K, Tarkka M, Pehkonen E, Kaukinen L, Honkonen El, Kaukinen S. Ischemic preconditioning has beneficial effects on left ventricular hemodynamic function after coronary artery bypass grafting operation. *Scand Cardiovasc J* 2000; 34: 242-53.
48. Wu Z-K, Pehkonen E, Laurikka J, Kaukinen L, Honkonen El, Kaukinen S, Laippala P, Tarkka MR. The protective effects of preconditioning decline in aged patients undergoing coronary artery bypass grafting. *J Thorac Cardiovasc Surg* 2001; 122: 972-8.
49. Wu Z-K, Livainen T, Pehkonen E, Laurikka J, Tarkka MR. Ischemic preconditioning suppresses ventricular tachyarrhythmias after myocardial revascularization. *Circulation* 2002; 106: 3091-6.
50. Wu Z-K, Livainen T, Pehkonen E, Laurikka J, Zhang S, Tarkka MR. Fibrillation in patients subjected to coronary artery bypass grafting. *J Thorac Cardiovasc Surg* 2003; 126: 1477-82.
51. Wu Z-K, Livainen T, Laurikka J, Saraste A, Kyto V, Pehkonen E, Savunen T, Tarkka MR. Cardiomyocyte apoptosis and ischemic preconditioning in open heart operations. *Ann Thorac Surg* 2001; 76: 528-34.

52. Wang X, Wei M, Kuukasjarvi P, Laurikka J, Jarvinen O, Rinne T, Honkonen El, Tarkka M. Novel pharmacological preconditioning with diaxoxide attenuates myocardial stunning in coronary artery bypass grafting. *Eur J Cardiothorac Surg* 2003; 24: 967-73.
53. Venugopal V, Hausenloy DJ, Ludman A, Di Salvo C, Kolvekar S, Yap J, Lawrence D, Bognolo J, Yellon DM. Remote Ischaemic preconditioning reduces myocardial injury in patients undergoing cardiac surgery with cold blood cardioplegia:a randomised controlled trial. *Heart* 2009; 95: 1567-71.
54. Zhao ZQ, Corvera JS, Halkos ME, Kerendi F, Wang NP, Guyton RA, Vinten-Johansen J. Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. *Am J Physiol* 2003; 285: H579-88.
55. Kin H, Zhao ZQ, Sun H-Y, Wang N-P, Corvera JS, Halkos ME, Kerendi F, Guyton RA, Vinten-Johansen J. Postconditioning attenuates myocardial ischemia-reperfusion injury by inhibiting events in the early minutes of reperfusion. *Cardiovasc Res* 2004; 62: 74-85.
56. Hearse DJ, Bolli R. Reperfusion induced injury: manifestations, mechanisms and clinical relevance. *Cardiovasc Res* 1992;256:H1719-25.
57. Zhang F, Chen G, Chen C, Yan M. Sevoflurane postconditioning converts persistent ventricular fibrillation into regular rhythm. *Eur J Anaesthesiol* 2009; 26: 766-71.
58. Sun K, Liu ZS, Sun Q. Role of mitochondria in cell apoptosis during hepatic ischemia-reperfusion injury and protective effect of ischemic postconditioning. *World J Gastroenterol* 2004; 10: 1934-8.
59. Zhao H, Sapolisky RM, Steinberg GK. Interrupting reperfusion as a stroke therapy: ischemic postconditioning reduces infarct size after focal ischemia in rats. *J Cereb Blood Flow Metab* 2006; 26: 1114-21.
60. Danielisová V, Gottlieb M, Némethová M, Burda J. Effects of Bradykinin Postconditioning on Endogenous Antioxidant Enzyme Activity After Transient Forebrain Ischemia in Rat. *Neurochem Res* 2008; 33: 1057-64.
61. Xing B, Chen H, Zhang M, Zhao D, Jiang R, Liu X, Zhang S. Ischemic postconditioning inhibits apoptosis after focal cerebral ischemia/reperfusion injury in the rat. *Stroke* 2008; 39:2362-9.
62. Liu X, Chen H, Zhan B, Xing B, Zhou J, Zhu H, Chen Z. Attenuation of reperfusion injury by renal ischemic postconditioning: The role of NO. *Biochem Biophys Res Commun* 2007; 359: 628-34.
63. Szwarc I, Soullier S, Gayrard N, Mejean C, Mourad G, Argiles A. Ischemic postconditioning prevents ischemic acute renal failure. *Transplant Proc* 2007; 39: 2554-6.
64. Serviddio G, Romano AD, Gesualdo L, Tamborra R, Di Palma AM, Rollo T, Altomare E, Vendemiale G. Nephrol Dial Transplant 2008; 23: 1504-12.
65. Zhang ZY, Liu XH, Guo XS, Liu FY. [Calreticulin is involved in ischemic postconditioning-induced attenuation of ischemia/reperfusion injury in rat skeletal muscle.] *Sheng Li Xue Bao* 2007; 59: 643-50. [Article in Chinese].
66. Moon JG, Lim HC, Gye MR, Oh JS, Park JW. Postconditioning attenuates ischemia-reperfusion injury in rat skin flap. *Microsurgery* 2008; 28: 531-7.
67. Şengül İ, Hasanoğlu A, Şengül D, Urhan MK, Taner AŞ, Güler O. Sonradan koşullanmanın; reperfüzyonun ilk dakikalarındaki olayların inhibisyonu yoluyla, intestinal iskemi-reperfüzyon hasarına etkisi. *Ulusal Cerrahi Kongresi* 2006, 24-28 Mayıs 2006 Antalya. Özeti Kitabı P-41.
68. Santos CH, Gomes OM, Pontes JC, Miji LN, Bispo MA. The ischemic preconditioning and postconditioning effect on the intestinal mucosa of rats undergoing mesenteric ischemia/reperfusion procedure. *Acta Cir Bras* 2008; 23: 22-8.
69. Liu KX, Li YS, Huang WQ, Chen SQ, Wang ZX, Liu JX, Xia Z. Immediate postconditioning during reperfusion attenuates intestinal injury. *Intensive Care Med*. 2009; 35: 933-42.
70. Dos Santos CH, Pontes JC, Gomes OM, Miji LN, Bispo MA. Evaluation of ischemic postconditioning effect on mesenteric ischemia treatment: experimental study in rats. *Rev Bras Cir Cardiovasc*. 2009; 24: 150-6.
71. Chu WW, Nie L, He XY, Yan AL, Zhou Y, Wu GL, Wang DH. [Change of cytochrome c in postconditioning attenuating ischemia-reperfusion-induced mucosal apoptosis in rat intestine.] *Sheng Li Xue Bao*. 2010;62:143-8. Chinese.
72. Garlid KD. Opening mitochondrial K-ATP in the heart - what happens and what does not happen. *Basic Res Cardiol* 2000; 95: 275-9.

73. Halkos ME, Kerendi F, Corvera JS, Wang NP, Payne CS, un HY, Guyton RA, Vinten-Johansen J, Zhao Z.Q. Myocardial protection with postconditioning is not enhanced by ischemic preconditioning. *Ann Thorac Surg* 2004; 78: 961-9.
74. Tsang A, Hausenloy DJ, Mocanu MM, Yellon DM. Postconditioning: a form of “modified reperfusion” protects myocardium by activating Phosphatidylinositol 3-kinase-Akt pathway. *Circ Res* 2004; 95: 230-2.
75. Yang X-M, Proctor JB, Cui L, Krieg T, Doney JM, Cohen MV. Multiple brief coronary occlusions during early reperfusion protects rabbit hearts by targeting cell signal pathways. *J Am Coll* 2004; 44: 1103-10.
76. Sun YH, Wang NP, Kerendi F, Halkos ME, Kin H, Guyton RA, Vinten-Johansen J, Zhao ZQ. Hypoxic postconditioning reduce cardiomyocyte loss by inhibiting ROS generation and intracellular Ca++ overload. *Am J Physiol Heart Circ Physiol* 2005; 4,1900-8.
77. Laskey WK. Brief repetitive balloon occlusion enhance reperfusion during percutaneous coronary intervention for acute myocardial infarction: a pilot study. *Catheter Cardiovasc Interv* 2005; 65: 361-7.
78. Staats P, Rioufol G, Piot C, Cottin Y, Cung TT, L'Huillier I, Aupetit JF, Bonnefoy E, Finet G, André-Fouët X, Ovize M. Postconditioning in the human heart. *Circulation* 2005; 112: 2143-8,
79. Thibault H, Piot C, Staats P, Bontemps L, Sportouch C, Rioufol G, Cung TT, Bonnefoy E, Angoulvant D, Aupetit JF, Finet G, André-Fouët X, Macia JC, Raczka F, Rossi R, Itti R, Kirkorian G, Derumeaux G, Ovize M. Long – term benefit of postconditioning. *Circulation* 2008; 117: 1037-44.
80. Luo W, Li B, Lin G, Huang R. Postconditioning in cardiac surgery for tetralogy of Fallot. *J Thorac Cardiovasc Surg* 2007; 133: 1373-4.
81. Luo W, Li B, Chen R, Huang R, Lin G. Effect of ischemic postconditioning in adult valve replacement. *Eur J Cardio - Thorac* 2008; 33: 203-8.
82. Li B, Chen R, Huang R, Luo W. Clinical benefit of cardiac ischemic postconditioning in corrections of tetralogy of Fallot. *Interact Cardio Vasc Thorac Surg* 2009; 8: 17-21.
83. Luo W, Li B, Lin G, Chen R, Huang R. Does cardioplegia leave room for postconditioning in paediatric cardiac surgery? *Cardiol Young* 2008; 18: 282-7.
84. Ramzy D, Rao V, Weisel RD. Clinical applicability of preconditioning and postconditioning: The cardiothoracic surgeon's view. *Cardovasc Res* 2006; 70: 174-80.
85. Venugopal V, Ludman A, Yellon DM, Hausenloy DJ. ‘Conditioning’ the heart during surgery. *Eur J Cardio-Thorac Surg* 2009; 35: 977-87.